

# General European OMCL Network (GEON)

## QUALITY MANAGEMENT DOCUMENT

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QUALIFICATION OF EQUIPMENT

QUALIFICATION OF ANALYTICAL COLUMNS

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## **ANNEX 11 OF THE OMCL NETWORK GUIDELINE “QUALIFICATION OF EQUIPMENT”**

### **QUALIFICATION OF ANALYTICAL COLUMNS**

*Note: Mandatory requirements in this guideline and its annexes are defined using the terms “shall” or “must”. The use of “should” indicates a recommendation. For these parts of the text other appropriately justified approaches are acceptable. The term “can” indicates a possibility or an example with non-binding character.*

#### **1. INTRODUCTION**

The present document outlines the general principles for the qualification of analytical columns used within OMCLs. Analytical columns are considered to be reagents, and as an essential part of the chromatographic system (Ph. Eur. Chapters 2.2.28, 2.2.29), they can influence the results obtained (ISO/IEC 17025:2017, Chapter 6.4 Equipment, point 6.4.1). For this reason, qualification of analytical columns should be carried out whenever considered necessary by the OMCL.

The OMCL is responsible for defining a suitable qualification scheme for use during the life cycle of the column, depending, for example, on the intended purpose and the frequency of use.

#### **2. SCOPE**

This document describes the requirements and general criteria for the qualification of analytical columns used in Liquid Chromatography (LC, UHPLC) and Gas Chromatography (GC) procedures.

It can be used as guidance by OMCLs when planning, performing and documenting the qualification process, upon receipt and during the life cycle of analytical columns.

Exemplary procedures provided in Annexes 1 and 2 have non-binding character. They can be helpful when carrying out the required qualification. Nevertheless, it is left to the professional judgment and background experience of each OMCL to apply the most suitable procedure, in order to prove that the analytical columns are suitable for their intended use.

#### **3. GLOSSARY AND DEFINITIONS**

The terminology used in this document is consistent with that used in Ph. Eur. Chapter 2.2.46 “Chromatographic separation techniques”.

#### **4. CONSIDERATIONS FOR QUALIFICATION UPON PURCHASING**

Incoming columns shall be verified for physical damage due to shipping, for compliance with the purchase order and for completeness of the documentation.

The OMCL shall retain records related to the management of columns for the applicable requirements described in ISO 17025:2017 clause 6.4.13.

#### **5. INITIAL QUALIFICATION**

The analytical column can be appropriately qualified before use by the OMCL according to a pre-defined procedure. This can be performed on receipt or, at the latest, prior to first use.

Initial column qualification, if carried out, should be documented in a suitable qualification record (including, e.g. date of qualification, reference to the procedure for qualification, reference materials or reagents used, results, chromatographic equipment used, pass/fail conclusion, etc.). This record can be used to make a complaint in cases where the column does not comply with the manufacturer's specification.

Verification of column performance can be done using one or more of the following approaches:

- a) by testing the column in the same conditions as given in the quality control leaflet provided by the manufacturer. The same parameters have to be checked and the results have to be compared with the manufacturer's results. Alternatively, other test mixtures may be used.
- b) If the column is used only for specific test methods, it might be appropriate to perform the qualification with the system suitability test described for the test method.
- c) Alternatively, columns with similar stationary phases can be qualified using a general test mixture/procedure.

### **Qualification parameters**

It is the responsibility of the OMCL to choose the most appropriate procedure, parameters and acceptance criteria. These should be appropriate to the intended use of the column. These parameters can include resolution, symmetry factor, theoretical plates, etc. Some typical limits are given as examples in Annexes 1 and 2.

## **6. PERIODICAL QUALIFICATION**

The qualification of the columns can be performed periodically using the same test compounds/procedure. A systematic record of the qualification results should be maintained in the laboratory.

### **Qualification procedure**

At least one of the procedures chosen for initial qualification can be applied for periodical qualification. The first chromatogram obtained with a new column in the laboratory can serve as a reference of column performance at the initial time point and used as a reference for subsequent verifications. Slight variations can be observed on chromatograms due to different LC or GC configuration of the equipment (e.g. dead volumes), operating environment, system electronics, etc. These can be taken into consideration whenever the acceptance criteria are set in-house.

### **Frequency of qualification**

The laboratory can define and document the frequency of qualification whenever periodic qualification is considered appropriate and useful. The frequency depends on the use of the column (e.g. routine or "spot" analysis). For instance, a qualification according to section 5 (above) could be performed at fixed time intervals (e.g. once a year) or depending on the extent of use, e.g. after a defined number of injections.

### **Qualification records**

A record of the qualification results can help the user to monitor the column performance over time and inform the decision to discard it. A column is considered suitable for the intended use when the acceptance/system suitability criteria are met.

## 7. IN-USE QUALIFICATION

In-use qualification corresponds to the system suitability test for the method used. It shall be performed on a regular basis before each test and/or during the sequence of injections.

The procedure (e.g. mobile phase) and the number of samples injected can contribute to the degradation of the stationary phase. This information should be traceable for each column, including, for example, the following information: samples analysed and matrix for finished products, chromatography conditions (mobile phase, temperature, pH value...), storage conditions, etc.

## 8. EXAMPLE PROCEDURES FOR QUALIFICATION OF ANALYTICAL COLUMNS

Most of the examples given below in Annexes 1 and 2 are qualification procedures provided by manufacturers.

The parameters tested for the compliance of the column performance qualification should be calculated and reported as defined by Ph. Eur. Chapter 2.2.46.

Consider any differences that might occur between the parameters defined in the manufacturer specification or instrument default calculations and the requirements of Ph. Eur. Chapter 2.2.46. (Note: the tailing factor calculated by the instrument software is quite often USP tailing by default).

## 9. REFERENCES

- 1) ISO/IEC 17025:2017 General requirements for the competence of testing and calibration laboratories.
- 2) Ph. Eur. Chapter 2.2.28. "Gas chromatography".
- 3) Ph. Eur. Chapter 2.2.29 "Liquid chromatography".
- 4) Ph. Eur. Chapter 2.2.46 "Chromatographic separation techniques".
- 5) Column performance test mixture for liquid chromatography - standard reference material 870. NIST.
- 6) HPLC column classification. USP Pharmacopeia Forum, Vol. 31(2).

**TABLE 1: EXAMPLES FOR QUALIFICATION OF ANALYTICAL COLUMNS FOR LC**

The chromatographic conditions provided below are applicable to a column size of 250 mm (length) × 4.6 mm (diameter). Where a column with other dimensions is used, the chromatographic conditions have to be adjusted according to Ph. Eur. chapter 2.2.46.

TYPE of COLUMN	Chromatographic settings:	Test mixture of: (solvent mobile phase)	Acceptance criteria
RP-LC C-8 and C-18 (also appropriate for C-4).	Mobile phase: acetonitrile:water = 58:42 v/v % Flow: 1.0 mL/min Detection: 254 nm Temperature: room temperature Injection volume: 5 µL	Uracil: 0.005 mg/mL Toluene: 4.0 mg/mL Phenol: 0.7 mg/mL N, N-diethyl-m- toluamide: 0.6 mg/mL	$A_s = 0.8$ to 1.5 $R_s > 1.5$ (between adjacent peaks) $N$ (toluene) > 3000 $\alpha_{\text{phenol/toluene}}$ and $\alpha_{\text{N,N-diethyl-m-toluamide/toluene}} \geq 1.5$ Uracil – indicator for the hold-up time ( $t_M$ )
RP-LC CN	Mobile phase: methanol:water = 70:30 v/v % Flow: 1.0 mL/min Detection: 254 nm Temperature: room temperature Injection volume: 5 µL	Uracil: 10 µg/mL Phenol: 200 µg/mL Toluene: 800 µg/mL 4-Cl-nitrobenzene: 25 µg/mL Naphthalene: 40 µg/mL	$A_s = 0.8$ to 1.5 $N$ (Naphthalene) > 3000 $R_s \geq 1.5$ (between adjacent peaks) Uracil – indicator for the hold-up time ( $t_M$ )
RP-LC PHENYL (phenyl propyl and phenyl hexyl)	Mobile phase: acetonitrile:water = 65:35 v/v % Flow: 1.0 mL/min Detection: 254 nm Temperature: room temperature Injection volume: 1 µL	Uracil: 0.01 mg/mL Acetophenone: 0.22 mg/mL Toluene: 9.42 mg/mL Naphthalene: 9.42 mg/mL	$A_s = 0.8$ to 1.5 $R_s > 1.5$ (between adjacent peaks) $N$ (Naphthalene) > 3000 $\alpha_{\text{acetophenone/toluene}}$ and $\alpha_{\text{toluene/naphthalene}} \geq 1.5$ Uracil – indicator for the hold-up time ( $t_M$ )
NP-LC (for Si, NH <sub>2</sub> , NO <sub>2</sub> , Diol, CN)	Mobile phase: hexane:ethanol R = 95:5 v/v % Flow: 1.0 mL/min Detection: 254 nm Temperature: room temperature Injection volume: 5 µL	Toluene: 1.0 mg/mL Diethyl phthalate: 1.0 mg/mL Dimethyl phthalate: 1.0 mg/mL	$A_s = 0.8$ to 1.5 $R_s > 1.5$ (between adjacent peaks) $N > 3000$ $\alpha_{\text{diethyl phthalate/dimethyl phthalate}} \geq 1.5$ Toluene – indicator for the hold-up time ( $t_M$ )
STRONG AND WEAK CATION-EXCHANGE (SCX; WCX)	Mobile phase: 0.15 M di- ammonium hydrogen phosphate buffer pH=6.0 Flow: 1.0 mL/min Detection: 254 nm Temperature: room temperature Injection volume: 10 µL	Uracil: 7 µg/mL Cytosine: 7 µg/mL (solvent: water)	$A_s = 0.8$ to 1.5 $N > 3000$ $R_s > 1.5$ $\alpha_{\text{uracil/cytosine}} \geq 1.5$

TYPE of COLUMN	Chromatographic settings:	Test mixture of: (solvent mobile phase)	Acceptance criteria
STRONG AND WEAK ANION-EXCHANGE COLUMNS (SAX; WAX)	Mobile phase: 0.05 M sodium di-hydrogen phosphate buffer pH=3.0 Flow: 1.0 mL/min Detection: 254 nm Temperature: room temperature Injection volume: 10 µL	Uridine: 7 µg/mL Uridine monophosphate: 7 µg/mL	$A_s = 0.8$ to 1.5 $N > 3000$ $R_s > 1.5$ $\alpha_{\text{uridine/uridine monophosphate}} \geq 1.5$
QUALIFICATION OF CHIRAL COLUMNS		It is recommended to test this column type with a test mixture according to the QC certificate of the manufacturer.	Results should be compared with those in the QC chromatogram of the manufacturer.
SIZE-EXCLUSION COLUMNS A very widely-used column in the quality control of therapeutic proteins (e.g. immunoglobulin monographs 0338 and 0918 and albumin monograph 0255) is based on hydrophilic silica gel for chromatography R with a fractionation range for protein of 10000 to 500000.	Mobile phase: dissolve 4.873 g disodium hydrogen phosphate dihydrate R, 1.741 g sodium dihydrogen phosphate monohydrate R and 11.688 g sodium chloride R in 1 L of water R.  Column dimensions: - 10 µm particle size: l = 600 mm, I.D.: 7.5 mm - 5 µm particle size: l = 300 mm, I.D.: 7.5 mm  Flow rate: 0.5 mL/min Detection: 280 nm Inj. Volume: 20 µL	Thyroglobulin (bovine, MW 670,000): 5.0 g/L γ-Globulin (bovine, MW 158,000): 5.0 g/L Ovalbumin (chicken, MW 44,000): 5.0 g/L Myoglobin (horse, MW 17,000): 2.5 g/L Vitamin B12 (MW 1,350): 0.5 g/L	The elution order is Thyroglobulin, γ-Globulin, Ovalbumin, Myoglobin and Vitamin B12. There may be additional peaks between Thyroglobulin and γ-Globulin (γ-Globulin-dimer peak) and in front of thyroglobulin (aggregates peak). $N$ (Vitamin B12): NLT 20,000 $A_s$ (Vitamin B12): 0.8 to 1.5 $R_s$ (Myoglobin/Ovalbumin): NLT 2.5

## ANNEX 2

TABLE 1: EXAMPLES FOR THE QUALIFICATION OF ANALYTICAL COLUMNS FOR GAS CHROMATOGRAPHY

TYPE of COLUMN	<i>Chromatographic settings</i> (FID detector):	<i>Test mixture of:</i> ( <i>solvent: methylene chloride</i> )	<i>Acceptance criteria</i>
NON-POLAR	<p><i>Narrow-bore columns (0.25 or 0.32 mm ID):</i> Carrier gas: Helium Linear gas rate: 20-25 cm/sec Injection port temperature: 220°C Oven temperature: 100-135°C FID temperature: 220°C Split ratio: 100:1 (5 ng of each component is delivered onto the column) Injection volume: 1 µL</p> <p><i>Wide-bore capillary columns (0.53 or 0.75 mm ID):</i> Dilute the mixture 1:10 in methylene chloride and inject 0.2 µL onto the column in direct injection mode.</p>	2-octanone: 0.5 mg/mL Decane: 0.5 mg/mL 1-octanol: 0.5 mg/mL 2,6-dimethyl-phenole: 0.5 mg/mL Undecane: 0.5 mg/mL 2,6-dimethylaniline: 0.5 mg/mL Dodecane: 0.5 mg/mL Tridecane: 0.5 mg/mL	$A_s = 0.8$ to 1.5 Plates/metre (peak with $k$ between 5 and 7): requirements given in Table2 (see below) $\alpha$ between adjacent peaks $\geq 1.5$ Methane –indicator for the hold-up time ( $t_M$ )
MEDIUM POLAR	<p><i>Narrow bore columns (0.25 or 0.32 mm ID):</i> Carrier gas: Helium Linear gas rate: 20-25 cm/sec Injection port temperature: 220°C Oven temperature: 100-135°C FID temperature: 220°C Split ratio: 100:1 (5 ng of each component is delivered onto the column) Injection volume: 1 µL</p> <p><i>Wide-bore capillary columns (0.53 or 0.75 mm ID):</i> Dilute the mixture 1:10 with methylene chloride and inject 0.2 µL onto the column in direct injection mode.</p>	N-decane: 0.5 mg/mL N-dodecane: 0.5 mg/mL N-tetradecane: 0.5 mg/mL N-tridecane: 0.5 mg/mL N-undecane: 0.5 mg/mL 1-octanol: 0.5 mg/mL 2-octanone: 0.5 mg/mL 2,6-dimethylaniline: 0.5 mg/mL 2,6-dimethyl-phenole: 0.5 mg/mL	$A_s = 0.8$ to 1.5 Plates/metre (peak with $k$ between 5 and 7): requirements given in Table 2 (see below) $\alpha$ between adjacent peaks $\geq 1.5$ Methane –indicator for the hold-up time ( $t_M$ )
POLAR	<p><i>Narrow-bore columns (0.25 or 0.32 mm ID):</i> Carrier gas: Helium Linear gas rate: 20-25 cm/sec Injection port temperature: 220°C Oven temperature: 145-185°C FID temperature: 220°C Split ratio: 100:1 (5 ng of each component is delivered onto the column) Injection volume: 1 µL</p> <p><i>Wide-bore capillary columns (0.53 or 0.75 mm ID):</i> Dilute the mixture</p>	N-eicosane: 0.5 mg/mL N-heptadecane: 0.5 mg/mL N-hexadecane: 0.5 mg/mL N-octadecane: 0.5 mg/mL N-pentadecane: 0.5 mg/mL 1-octanol: 0.5 mg/mL 2-octanone: 0.5 mg/mL 2,6-dimethylaniline: 0.5 mg/mL 2,6-dimethyl-phenole: 0.5 mg/mL	$A_s = 0.8$ to 1.5 Plates/metre (peak with $k$ between 5 and 7): requirements given in Table 2 (see below) $\alpha$ between adjacent peaks $\geq 1.5$ Methane –indicator for the hold-up time ( $t_M$ )

TYPE of COLUMN	Chromatographic settings (FID detector):	Test mixture of: (solvent: methylene chloride)	Acceptance criteria
	1:10 with methylene chloride and inject 0.2 $\mu$ L onto the column in direct injection mode.		

**LEGEND:**A<sub>s</sub>: Symmetry factorR<sub>s</sub>: Resolution

N: Plate number

 $\alpha$ : selectivity factor ( $\alpha$ ) i.e. ratio of the retention factors (k) of the two adjacent peaks**Plate number**

The column performance may be calculated for a given component as the plate number (also referred to as number of theoretical plates).

This parameter is very useful for comparisons of GC columns. Table 2 below represents the requirements for the average column performance, given in plates/metre, according to the internal diameter and polarity of the phase.

*Table 2. Comparison of average efficiency by phase polarity and internal diameter*

Internal diameter	Polarity of the column		
	Non-polar	Intermediate polarity	Polar
0.1 mm	10000	-	-
0.2 mm	4500	4200	4000
0.32 mm	3200	3000	2500
0.53 mm	1500	1350	1300